



ISSN: 0974 - 0376

The Ecoscan : Special issue, Vol. VIII: 241-246: 2015
AN INTERNATIONAL QUARTERLY JOURNAL OF ENVIRONMENTAL SCIENCES
www.theecoscan.in

PROLINE ACCUMULATION FOR IMPROVING STAY GREEN TRAIT TO COMBAT WITH ENVIRONMENTAL ABIOTIC STRESS IN WHEAT (*TRITICUM AESTIVUM*)

Naresh Pratap Singh and Vaishali

KEYWORDS

Ethyl Methanesulphonate
Stay green
Biochemical
Proline
Mutated Wheat

**Proceedings of National Conference on
Harmony with Nature in Context of
Bioresources and Environmental Health
(HARMONY - 2015)**
November 23 - 25, 2015, Aurangabad,
organized by
Department of Zoology,
Dr. Babasaheb Ambedkar Marathwada University
Aurangabad (Maharashtra) 431 004
in association with
National Environmentalists Association, India
www.neaindia.org



NARESH PRATAP SINGH* AND VAISHALI

Department of Biotechnology,
SVP University of Agriculture and Technology, Meerut (U.P.) - 250 110, INDIA
e-mail: naresh.singh55@yahoo.com

ABSTRACT

Proline accumulation is an important mechanism for osmotic regulation under abiotic stress. In this study, we evaluated proline accumulation in wheat (*Triticum aestivum*) varieties after treatment with Ethyl Methanesulphonate (EMS) of two concentrations (0.5% and 1.0%) for 90 minutes. The proline content was observed at booting and maturity stage. At booting stage, proline content was varied from 1.52 $\mu\text{mol/gfw}$ in K 7410 to 1.05 $\mu\text{mol/gfw}$ in variety HD 2177 in control plants. Variety K 7410 showed highest proline content ranges from 1.52 to 1.65 $\mu\text{mol/gfw}$ after the two treatments of EMS. At maturity stage, the proline content varied from 1.09 to 2.22 $\mu\text{mol/gfw}$ in control plants, 1.04 to 2.35 $\mu\text{mol/gfw}$ after treatment one (0.5%) and 1.00 to 2.46 $\mu\text{mol/gfw}$ after treatment second (1%) of EMS. Whereas, variety HD 2177 and K68 shows significant decreased level of proline content after two treatments i.e. 1.27 to 1.08 $\mu\text{mol/gfw}$ and 1.48 to 1.01 $\mu\text{mol/gfw}$. On the whole, these ten wheat varieties showed decreased level of proline after two treatments of EMS except K 7410 at both the stages. Such results indicate that EMS mutagen had changed some genetic alteration which led to high proline accumulation in wheat varieties which may perform better under abiotic stress conditions such as drought, salt, high temperature etc.

INTRODUCTION

Growth and productivity of major cereal crops like wheat, rice, maize etc. is greatly affected by environmental stresses such as drought, high salinity, and low temperature due to their sessile nature (Boyer, 1982; Eriksson *et al.*, 2011). Abiotic stress is also known as Osmotic stress which includes stomatal closure, decrease in photosynthesis and growth inhibition, accumulation of proline, mannitol etc. (Nagar *et al.*, 2015). So, to survive in such stressed environment, plants have developed a number of physiological and biochemical mechanisms (Bartels and Salamini, 2001; Wang *et al.*, 2003). These mechanisms initiate the expression of a variety of genes against the abiotic stresses by plants. Such genes help in protecting cells from stress by producing important metabolic proteins such as key enzymes for osmolyte biosynthesis, mainly proline, water channel proteins, sugar and proline transporters, chlorophyll *a/b* binding proteins (Fowler *et al.*, 2002; Kreps *et al.*, 2002; Seki *et al.*, 2002). Stay-green character, has been identified as an important component in the genetic improvement of several crops to promote stress tolerance and yield gain (Peigao *et al.*, 2006). Stay green phenotypes maintain green leaf area (chlorophyll) for a longer period (photosynthesis), delay leaf senescence which helps the plant to perform better in abiotic stress conditions (Spano *et al.*, 2003; Luchee *et al.*, 2015). Crops grown in stress environments as their yield is associated with the capacity of the plant to maintain CO_2 assimilation (Hafsiet *et al.*, 2007). Thus, this vital trait should be incorporated by developing genes for it using mutagens etc. into major crops like wheat, rice etc. Thomas and Howarth (2000) described five types of stay-green phenotypes: Type A phenotypes show late initiation of senescence with a normal senescence rate. Type B phenotypes show normal initiation of senescence with a slower rate of senescence. Type C phenotype shows lesion in chlorophyll degradation, leaving the rest of the senescence process unaffected. Type D phenotype shows rapid death (freeze, boil, dry) ensures maintenance of leaf colour in dead leaf. Type E: enhanced greenness but unchanged initiation and rate of senescence. As a result the overall process of senescence will take longer to complete. Types A, B and possibly E are functionally stay-green: they maintain photosynthetic capacity in their green tissues. Therefore, they may be a potential means to improve grain yield. In China wheat lines with a wheat-rye chromosome translocation were developed which showed a functional stay-green phenotype combined with increased grain yield and total biomass of up to 25% when grown in the field (Chen *et al.*, 2010; Luo *et al.*, 2006). Under abiotic stress conditions not only the stay green trait will help but also the accumulation of important metabolic proteins known as "compatible osmolytes" mainly proline, will help the plant to overcome the abiotic stress. The stay-green trait has been reported to increase yields (Gong *et al.*, 2005; Christopher *et al.*, 2008; Chen *et al.*, 2010), and there were positive correlations to water use efficiency, nitrogen use, leaf senescence with chlorophyll content (Górny and Garczyński, 2002; Kipp *et al.*, 2013; Gwendolin *et al.*, 2015), yields under heat and drought (Naruoka *et al.*, 2012). Many stay green mutants, referred to as functional stay green or non-yellowing in various plant species have been reported to maintain leaf greenness after the grain-ripening stage and give better yield (Spano *et al.*, 2003; Walulu *et al.*, 1994). In wheat, the

*Corresponding author

maintenance of proteins embedded in the walls of thylakoids and low levels of reactive oxygen species demonstrated the superiority of wheat mutant *tasg1* tolerant to drought (Tian et al., 2013). The stress tolerance varieties of wheat screened by breeding method (Arya et al., 2013) and may be studied at molecular level for stay green trait and proline. Therefore, the present research work has aimed to bring light to major aspects of the stay-green trait and proline for high biomass yield of crops as they both collectively play vital role in maintaining chlorophyll content, photosynthesis duration and osmotic potential under stressed condition.

MATERIALS AND METHODS

The present research work was carried out at laboratory of Department of Biotechnology, College of Agriculture, of S.V.P. University of Agriculture and Technology, Meerut during rabi season. A total of ten varieties of wheat (*Triticum aestivum*) cultivars viz. HUW 510, C 306, Sonalika, HD 2135, HD 2177, VL 401, K 9162, RAJ 3765, K 68, K 7410 were collected to study the effect of EMS treatment on biochemical character i.e. proline content. The seeds of all ten varieties were treated with EMS (0.5% and 1% in distilled water) for 90 minutes in petri plates with three replicates. Thereafter, the treated seeds of wheat were sown in pots and the proline content is observed from three replicates two times one at booting stage and second at final maturity stage of control and treatments.

Biochemical Characterization

Proline determination proceeded according to Bates et al., 1973 based on proline’s reaction with ninhydrin. For estimation of Proline content in wheat germplasm, 100 mg of fresh leaf tissue was taken from plants. Grind the leaf tissue in aqueous sulphosalicylic acid (3%). Centrifuged the tubes at 7000 rpm for 5 min. The supernatant was mixed with equal volume of Glacial acetic acid. Then add 0.5 ml of ninhydrin and incubated for 30 minute at 100 in boiling water bath and then placed for 5 min in ice bath for cooling. Thereafter 2 ml of toluene was mixed in reaction mixture and mixed properly. The aqueous phase was transferred in new tube. The reaction mixture was warm at 25°C and chromophore was measured at 520nm was determined in a BioMate spectrophotometer (ThermoSpectronic).

Statistical analysis

The experimental data obtained from randomly selected five plant from each replicates were subjected to the statistical analysis outlined by Panse and Sukhatme (1978). The significance of differences among treatment means was tested by ‘F’ test and critical differentiation (at 5 per cent probability) was calculated by the method given by Pearson K., 1895.

RESULTS AND DISCUSSION

Proline is an important parameter to know the tolerance capacity of the plants in stress conditions (Delauney and Verma, 1993), and its accumulation indicates the response against abiotic stresses like drought, salt, high temperature etc. (Ramanjulu and Sudhakar, 2000). Among the studied wheat (*Triticum aestivum*) varieties the proline content at booting stage in the leaf varied from 1.52 μmol/gfw in K 7410 to 1.05 μmol/gfw in variety HD 2177 in control plants (Table 1). All the variety showed a little decrement in the amount of total proline after the treatments of EMS except in Sonalika, K 7410 and RAJ 3765 (Figure 1). The highest proline content was shown by variety K 7410 and it found to be increases from 1.52 to 1.65 μmol/gfw after the two treatments of EMS. Variety Sonalika and RAJ 3765 also shown increment in proline content from 1.21 to 1.29 μmol/gfw and 1.49 to 1.55 μmol/gfw. Whereas, the variety HD 2177 showed the lowest amount of proline content and found to be decreases from 1.05 to 0.99 μmol/gfw after all two treatments. The similar pattern of increment of proline content was also observed at maturity stage in present investigation. The proline content varied from 1.09 to 2.22 μmol/gfw in control plants (Table 2). The amount of total proline content after the treatments of EMS in Sonalika, K 7410 and RAJ 3765 shows an increment level among from all wheat varieties. The wheat varieties other than Sonalika, K 7410 and RAJ 3765 shows slight decrease in proline level after two treatments but variety HD 2177 and K68 shows significant decreased level of proline content after all treatments 1.27 to 1.08 μmol/gfw and 1.48 to 1.01 μmol/gfw (Figure 2). The highest proline accumulation was shown by variety K 7410 after all treatments of EMS from 2.22 to 2.46 μmol/gfw. High proline synthesis in plants could favor a better recovery of plants in stressed conditions, converted in grain yield to a

Table 1: Proline content of ten wheat varieties of control and two treatments of EMS (at booting stage)

S.No.	Variety	Proline (μmol/gfw)		T1		T2	
		Control Mean	SE	Mean	SE	Mean	SE
1	HUW 510	1.21	±0.019	1.20	±0.006	1.18	±0.006
2	C 306	1.12	±0.043	1.05	±0.018	1.04	±0.015
3	Sonalika	1.21	±0.012	1.27	±0.025	1.29	±0.019
4	HD 2135	1.30	±0.009	1.28	±0.015	1.25	±0.009
5	HD 2177	1.05	±0.007	1.00	±0.012	1.00	±0.006
6	VL 401	1.40	±0.006	1.38	±0.021	1.33	±0.031
7	K 9162	1.08	±0.012	1.06	±0.012	1.04	±0.012
8	RAJ 3765	1.49	±0.013	1.52	±0.017	1.55	±0.012
9	K 68	1.28	±0.022	1.11	±0.007	1.10	±0.007
10	K 7410	1.52	±0.015	1.57	±0.009	1.65	±0.015
	S.Em ±			0.214			
	C.D. (p=0.05)			0.073			

T1 = 0.5% EMS, T2 = 1.0% EMS

Table 2: Proline content of ten wheat varieties of control and two treatments of EMS (at maturity stage)

S.No.	Variety	Proline ($\mu\text{mol/gfw}$)		T1		T2	
		Control Mean	SE	Mean	SE	Mean	SE
1	HUW 510	1.51	± 0.020	1.55	± 0.009	1.53	± 0.004
2	C 306	1.32	± 0.053	1.25	± 0.020	1.20	± 0.017
3	Sonalika	1.57	± 0.022	1.59	± 0.029	1.61	± 0.009
4	HD 2135	1.32	± 0.019	1.26	± 0.019	1.21	± 0.009
5	HD 2177	1.27	± 0.017	1.11	± 0.018	1.08	± 0.008
6	VL 401	1.50	± 0.016	1.45	± 0.027	1.40	± 0.038
7	K 9162	1.09	± 0.011	1.04	± 0.015	1.00	± 0.014
8	RAJ 3765	2.09	± 0.018	2.19	± 0.016	2.25	± 0.011
9	K 68	1.48	± 0.029	1.11	± 0.005	1.01	± 0.009
10	K 7410	2.22	± 0.012	2.35	± 0.010	2.46	± 0.011
	S.Em \pm			0.321			
	C.D. ($p=0.05$)			0.123			

T1 = 0.5% EMS, T2 = 1.0% EMS

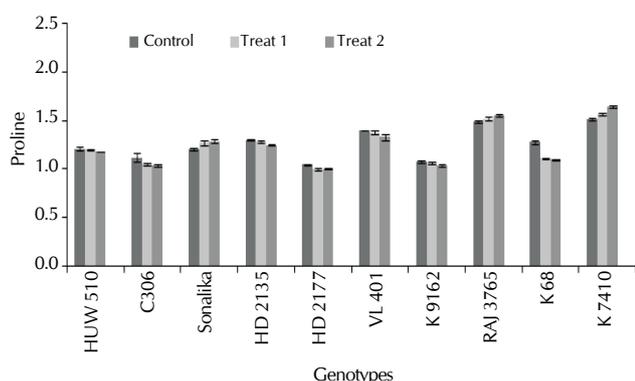


Figure 1: Graph representing the proline content in wheat at booting stage

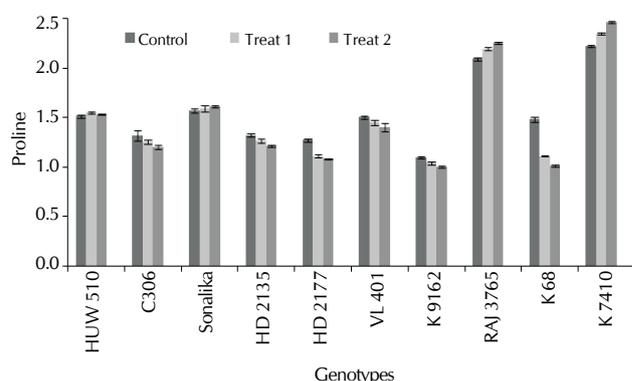


Figure 2: Graph representing the proline content in wheat at maturity stage

protected photosynthetic apparatus after water field status recovery. The similar results of proline levels increase in wheat under stress condition pointed by Nayyar (2003) and Zhu *et al.* (2005). Similarly, results are in agreement with those obtained by Chen and Li (2002), who showed that a high concentration of proline in suspension cells avoided lipid peroxidation (Parvanova *et al.*, 2004). Numerous studies have linked the accumulation of proline to abiotic stress (Aziz *et al.*, 1999; Munns and Tester, 2008; Ribeiro *et al.*, 2014), and it can play a protective role against the osmotic potential, membrane instability generated by salt, heat etc. (Chen *et al.*, 2007; Hoque *et al.*, 2008; Satbhai *et al.*, 2014). Proline accumulation under drought stress has been reported in other crop also like chick pea (Ayerb and Tenori, 1998), corn (Serraj and Sinclair, 2002) and peanut (Smith *et al.*, 2002). Huang *et al.* (2013) had determined the relationship between salt tolerance and proline content in Jerusalem artichoke plantlets. Synthesis or accumulation of proline is depending on the activities of enzymes such as pyrroline-5-carboxylate synthetase (P-5-C synthetase), pyrroline 5-carboxylate reductase etc. Several investigators (Ramanjulu and Sudhakar, 2000; Kumar *et al.*, 2003) reported a positive correlation between the accumulation of proline and its osmoprotective role at the whole plant level during abiotic stress conditions mainly in drought, salt etc. Similarly, in the present study a positive correlation between free proline accumulation and stay green

trait in wheat cultivars in control and treated plants which can adopt better themselves in abiotic stress conditions. Stay green trait help in maintaining the long photosynthetic duration and chlorophyll content which results better yield (Christopher *et al.*, 2008; Chen *et al.*, 2010). Total flag leaf photosynthesis, chlorophyll content, the onset of senescence and green leaf duration have all been found to be positively correlated with wheat grain yield (Kichey *et al.*, 2007; Wang *et al.*, 2008; Gajuet *et al.*, 2011). Hui *et al.* (2012) reported the generation of wheat mutant *tasgl*, induced using EMS exhibits delay in leaf senescence in term of chlorophyll. Four or five classes of delayed senescence or 'stay-green' have been described in the literature (Thomas and Smart, 1993; Thomas and Howarth, 2000) and the stay-green phenotype has shown proven utility to improve yields under abiotic stress (Harris *et al.*, 2007; Vijayalakshmi *et al.*, 2010) and also under biotic stress, like spot blotch (Joshi *et al.*, 2007). High level of Proline could increase the concentration of cell protoplasm, to maintain normal membrane function at high temperatures to be conducive to plant resistance of environmental stress, result proven in tomatoes by Yin *et al.* (2001). The present investigation further supports the stress tolerance of the Sonalika, K 7410 and RAJ 3765 based on greater maintenance of proline content coupled with stay green trait i.e. long duration leaf chlorophyll stability and photosynthesis after treatment of EMS. The study of stay-green character can

be beneficial in breeding programs, which helps in significant genetic progress for attributes such as high yield, industrial quality, disease resistance and tolerance to abiotic stresses. The understanding of the proline accumulation mechanism associated with leaf senescence and photosynthetic efficiency in several major crops may be the key to delete the plateau of productivity associated with adaptation to unfavorable environmental conditions.

REFERENCES

- Arya, S., Mishra, D. K. and Bornare, S. S. 2013. Screening genetic variability in advance lines for drought tolerance of bread wheat (*Triticumaestivum*). *The Bioscan*. **8(4)**: 1193-1196.
- Ayerbe, L. and Tenorio, J. L. 1998. Turgor maintenance, osmotic adjustment and soluble sugar and proline accumulation in 49 pea cultivars in response to water stress; *Field crop Res.* **59**: 225-235.
- Aziz, A., Martin-Tanguy, J. and Larher, F. 1999. Salt stress-induced proline accumulation and changes in tyramine and polyamine levels are linked to ionic adjustment in tomato leaf discs. *Plant Science*. **145**: 83-91.
- Bartels, D. and Salamini, F. 2001. Desiccation tolerance in the resurrection plant *Craterostigma plantagineum* L. A contribution to the study of drought tolerance at the molecular level; *Plant Physiology*. **127**: 1346-1353.
- Bates, L. S., Waldren, R. P. and Teare, I. D. 1973. Rapid determination of free proline for water-stress studies; *Plant and Soil*. **39**: 205-207.
- Boyer, J. S. 1982. Plant productivity and environment; *Science*. **218**: 443-448.
- Chen, J. B., Liang, Y., Hu, X. Y., Wang, X. X., Tan, F. Q., Zhang, H. Q., Ren, Z. L. and Luo, P. G. 2010. Physiological characterization of 'stay green' wheat cultivars during the grain filling stage under field growing conditions; *Acta. Physiol. Plant.* **32**: 875-882.
- Chen, W. and Li, P. H. 2002. Membrane stabilization by abscisic acid under cold aids proline in alleviating chilling injury in maize (*Zea mays* L.) cultured cells; *Plant Cell Environ.* **25**: 955-62.
- Chen, Z., Cuin, T. A., Zhou, M., Twomey, A., Naidu, B. P. and Shabala, S. 2007. Compatible solute accumulation and stressmitigating effects in barley genotypes contrasting in their salt tolerance. *J. Experimental Botany*. **58**: 4245-4255.
- Christopher, J. T., Manschadi, A. M., Hammer, G. L. and Borrell, A.K. 2008. Developmental and physiological traits associated with high yield and stay-green phenotype in wheat; *Australian J. Agricultural Research*. **59**: 354-364.
- Delauney, A. J. and Verma, D. P. S. 1993. Proline biosynthesis and osmoregulation in plants; *The Plant J.* **4**: 215-223.
- Eriksson, S. K. and Harryson, P. 2011. Dehydrins: molecular biology, structure and function; *Plant Desiccation Tolerance*, (eds Lutte et al., Springer verlag). pp. 289-305
- Flowers, T. J. and Yeo, A. R. 1995. Breeding for salinity resistance in crop plants; *Aust. J. Plant Physiol.* **22**: 875-884.
- Gaju, O., Allard, V., Martre, P., Snape, J.W., Heumez, E., Gouis, J., Moreau, D., Bogard, M., Griffiths, S., Orford, S., Hubbard, S. and Foulkes, M. J. 2011. Identification of traits to improve the nitrogen-use efficiency of wheat genotypes. *Field Crops Research*. **123**: 139-152.
- G'orny, A. G. and Garczy'nski, S. 2002. Genotypic and nutrition-dependent variation in water use efficiency and photosynthetic activity of leaves in winter wheat (*Triticumaestivum* L.); *J. Appl. Genet.* **43**: 145-160.
- Gong, Y. H., Zhang, J., Gao, J. F., Lu, J. Y. and Wang, J. R. 2005. Slow export of photoassimilate from stay-green leaves during late grain-filling stage in hybrid winter wheat (*Triticumaestivum*L.); *J. Agronomy and Crop Science*. **191**: 292-299.
- Gwendolin, G. W., Christiane, C. B., Matthias, M. E., Klaus, K. H. and Frank, F. O. 2015. Identification of genomic regions involved in tolerance to drought stress and drought stress induced leaf senescence in juvenile barley; *BMC Plant Biology*. **15**:125.
- Hafsi, M., Akhter, J. and Monneveux, P. 2007. Leaf senescence and carbon isotope discrimination in durum wheat (*Triticum durum* desf.) under severe drought conditions; *Cereal Research Communications*. **35**: 71-80.
- Harris, K., Subudhi, P.K., Borrel, A., Jordan, D., Rosenow, D., Nguyen, H., Klein, P., Klein, R. and Mullet, J. 2007. Sorghum stay-green QTL individually reduce post-flowering drought-induced leaf senescence; *J. Experimental Botany*. **58**: 327-338.
- Hoque, M. A., Banu, M. N., Nakamura, Y., Shimoishi, Y. and Murata, Y. 2008. Proline and glycinebetaine enhance antioxidant defense and methylglyoxal detoxification systems and reduce NaCl-induced damage in cultured tobacco cells; *J. Plant Physiology*. **165**: 813-824.
- Huang, Z., Zhao, L., Chen, D., Liang, M., Liu, Z., Shao, H. and Xiaohua Long, X. 2013. Salt Stress Encourages Proline Accumulation by Regulating Proline Biosynthesis and Degradation in Jerusalem Artichoke Plantlets; *PLoS ONE*. **8(4)**: e62085.
- Hui, Z., Tian, F. X., Wang, G. K., Wang, G. P. and Wang, W. 2012. The antioxidativedefence system is involved in the delayed senescence in a wheat mutagen *tasgl*; *Plant Cell Rep.* **31**: 1073-1084.
- Joshi, A. K., Kumari, M., Singh, V. P., Reddy, C. M. Kumar, S., Rane, J. and Chand, R. 2007. Stay green trait: variation, inheritance and its association with spot blotch resistance in spring wheat (*Triticumaestivum* L.); *Euphytica*. **153**: 59-71.
- Kichey, T., Hirel, B., Heumez, E., Dubois, F. and Gouis, J. 2007. In winter wheat (*Triticumaestivum* L.), post-anthesis nitrogen uptake and remobilisation to the grain correlates with agronomic traits and nitrogen physiological markers; *Field Crops Res.* **102**: 22-32.
- Kipp, S., Bodo, M. and Schmidhalter, U. 2013. Identification of stay-green and early senescence phenotypes in high-yielding winter wheat, and their relationship to grain yield and grain protein concentration using high-throughput phenotyping techniques, *Functional Plant Biology*. **41(3)**: 227-235.
- Kreps, J. A., Wu, Y., Chang, H. S., Zhu, T., Wang, X. and Harper, J. F. 2002. Transcriptome changes for Arabidopsis in response to salt, osmotic, and cold stress; *Plant Physiol.* **130**: 2129-2141.
- Kumar, S. G., Reddy, A. M. and Sudhakar, C. 2003. NaCl effects on proline metabolism in two high yielding genotypes of mulberry (*Morus alba* L.) with contrasting salt tolerance; *Plant Science*. **165**: 1245-1251.
- Lucho, H. S., Silva, J. A. G., Maia, L. C. and Oliveira, A. C. 2015. Stay-green: a potentiality in plant breeding; *Ciência Rural, Santa Maria*, **45(10)**: 1755-1760.
- Luo, P. G., Ren, Z. L., Wu, X. H., Zhang, H. Y., Zhang, H. Q. and Feng, J. 2006. Structural and biochemical mechanism responsible for stay green phenotype in common wheat; *Chin. Sci. Bull.* **51**: 2595-2603.
- Munns, R. and Tester, M. 2008. Mechanisms of salt tolerance; *Annual Review Plant Biology*. **59**: 651-681.
- Nagar, S., Aroral, A., Singh, V. P., Ramakrishnan, S., Umesh, D. K., Kumar, S. and Saini, R. P. 2015. Effect of cytokinin analogues on cytokinin metabolism and stress responsive genes under osmotic stress in wheat; *The Bioscan*. **10(1)**: 67-72.
- Naruoka, Y., Sherman, J. D., Lanning, S. P., Blake, N. K., Martin, J. M. and Talbert, L. E. 2012. Genetic analysis of green leaf duration in spring wheat. *Crop Sci.* **52**: 99-109.

- Nayyar, H. 2003.** Accumulation of osmolytes and osmotic adjustment in water-stressed wheat (*Triticum aestivum* L.) and maize (*Zea mays*) affected by calcium and its antagonists; *Environ Exp Bot.* **50**: 253-64.
- Panse, V. G. and Sukhatme, P. V. 1978.** Statistical methods for agricultural workers ICAR, New Delhi. pp. 145-150.
- Parvanova, D., Ivanov, S., Konstantinova, T., Karanov, E., Atanassov, A. and Tsvetkov, T. 2004.** Transgenic tobacco plants accumulating osmolytes show reduced oxidative damage under freezing stress; *Plant Physiol Biochem.* **42**: 57-63.
- Pearson, K. 1895.** Notes on regression and inheritance in the case of two parents; *Proceeding of the Royal Society of London.* **58**: 240-242.
- Peigao, L., Zhenglong, R., Xianhua, W. U., Huaiyu Z., Huaiqiong Z. and Juan, F. 2006.** Structural and biochemical mechanism responsible for the stay-green phenotype in common wheat; *Chinese Science Bulletin.* **51(21)**: 2595-2603.
- Ramanjulu, S. and Sudhakar, C. 2000.** Proline metabolism during dehydration in two mulberry genotypes with contrasting drought tolerance; *J. Plant Physiology.* **157**: 81-85.
- Ribeiro, R. C., Matias, J. R., Pelacani, C. R. and Dantas, B. 2014.** Activity of antioxidant enzymes and proline accumulation in *Erythrina velutina* Willd. seeds subjected to abiotic stresses during germination; *França J. Seed Science.* **36(2)**: 231-239.
- Satbhai, R., Kale, A. and Naik, R. 2014.** Cell viability, time laps study and membrane stability index during temperature induction response in wheat; *The Ecoscan.* **4**: 245-252.
- Seki, M., Narusaka, M., Ishida, J., Nanjo, T., Fujita, M., Oono, Y., Kamiya, A., Nakajima, M., Enju, A. and Sakurai, T. 2002.** Monitoring the expression profiles of ca 7000 *Arabidopsis* genes under drought, cold and high-salinity stresses using a full-length cDNA microarray; *Plant J.* **31**: 279-292.
- Serraj, R. and Sinclair, T. R. 2002.** Osmolyte accumulation: can it really help increase crop yield under drought condition; *Plant cell Environ.* **25**: 333-341.
- Smith, B.N., Girija, C. and Swamy, P.M. 2002.** Interactive effects of sodium chloride and calcium chloride on the accumulation of proline and glycine betaine in peanut (*Arachis hypogaea* L.); *Environ. and Exp. Bot.* **47**: 1-10.
- Spano, G., Fonzo, N. Di., Perrotta, C., Platani, C., Ronga, G., Lawlor, D. W., Napier, J. A. and Shewry, P. R. 2003.** Physiological characterization of 'stay green' mutants in durum wheat. *J. Experimental Botany.* **54**: 386.
- Thomas, H. and Smart, C. M. 1993.** Crops that stay green; *Ann. Appl. Biol.* **123**: 193-219.
- Thomas, H. and Howarth, C. J. 2000.** Five ways to stay green; *J. Exp. Bot.* **51**: 329-337.
- Tian, F., Gong, J., Zhang, J., Zhang, M., Wang, G., Li, A. and Wang, W. 2013.** Enhanced stability of thylakoid membrane proteins and antioxidant competence contribute to drought stress resistance in the *tag1* wheat stay-green mutant; *J. Experimental Botany.* **64(6)**: 1509-1520.
- Vijayalakshmi, K., Fritz, A. K., Paulsen, G. M., Bai, G. Pandravada, S. and Gill, B. S. 2010.** Modeling and mapping QTL for senescence-related traits in winter wheat under high temperature. *Molecular Breeding.* **26**: 163-175.
- Walulu, R. S., Rosenow, D. T., Wester, D. B. and Nguyen, H. T. 1994.** Inheritance of the stay-green trait in sorghum; *Crop Sci.* **34**: 970-972.
- Wang, H., McCaig, T.N., DePauw, R.M. and Clarke, J.M. 2008.** Flag leaf physiological traits in two high-yielding Canada Western Red Spring wheat cultivars. *Can. J. Plant Sci.* **88**: 35-42.
- Wang, W., Vinocur, B. and Altman, A. 2003.** Plant responses to drought, salinity and extreme temperatures: towards genetic engineering for stress tolerance; *Planta.* **218**: 1-14.
- Yin, X. G., Luo, Q. X. and Wang, W. Q. 2001.** Studies oil methodology for identification of heat tolerance of tomato; *Southwest China J. Agricultural Science.* **14**: 62-65.
- Zhu, X., Gong, H., Chen, G., Wang, S. and Zhang, C. 2005.** Different solute levels in two spring wheat cultivars induced by progressive field water stress at different developmental stages; *J. Arid. Environ.*, **62**: 1-14.