



ISSN: 0974 - 0376

The Ecoscan : Special issue, Vol. VII: 00-00: 2015
AN INTERNATIONAL QUARTERLY JOURNAL OF ENVIRONMENTAL SCIENCES
www.theecoscan.in

EFFECT OF ORGANIC, INORGANIC AND BIO-FERTILIZERS ON PRODUCTION OF LOOSE FLOWER-A REVIEW

Mridubhashini Patanwar *et al.*,

KEYWORDS

Bio-fertilizers
Flowering
Growth
Inorganic fertilizer
Loose flower
Organic fertilizer

**Proceedings of National Conference on
Harmony with Nature in Context of
Bioresources and Environmental Health
(HARMONY - 2015)**
November 23 - 25, 2015, Aurangabad,
organized by
Department of Zoology,
Dr. Babasaheb Ambedkar Marathwada University
Aurangabad (Maharashtra) 431 004
in association with
NATIONAL ENVIRONMENTALISTS ASSOCIATION, INDIA
www.neaindia.org



MRIDUBHASHINI PATANWAR*¹, PRITEE AWASTHY² AND BHAVESH PATEL¹

¹Department of Horticulture, Indira Gandhi Agriculture University, Raipur - 492 012, Chhattisgarh, INDIA

²Department of Agronomy, Indira Gandhi Agriculture University, Raipur - 492 012, Chhattisgarh, INDIA

e-mail: to.mridu09@gmail.com

ABSTRACT

Loose flowers are most popular and preferred by most of the farmer in India and Chhattisgarh. In India it occupied about 255 ('000 ha) with average production 2297 ('000 MT). Indian floriculture industry is very dynamic in its varieties and trade volumes demonstrating 6-9% of Annual growth. The effects of integrated use of organics with bio-fertilizers and inorganic fertilizers and impacts of long-term use of INM on enhancing crop productivity were studied on flowers. INM enhances the yield potential of crops over and above achievable yield with recommended fertilizers and results in better synchrony of crop N needs due to slower mineralization of organics, reduced N losses via denitrification, increase nitrogen fixation, phosphorous availability, lowering nitrate leaching, enhanced nutrient use efficiency, improvements in soil health and productivity and hence could sustain high crop yields in various cropping systems ensuring long-term sustainability of the system. In addition, this review there is greater opportunity to researcher to develop a perfect combination of organic, inorganic and biofertilizer for better production of loose flower.

INTRODUCTION

In a world that is increasingly becoming urbanized, ornamental plants provide an important link with the natural world. Karnataka, Tamil Nadu, Andhra Pradesh, West Bengal, Maharashtra, Uttarakhand, Uttar Pradesh, Delhi, Haryana, Kerala, Himachal Pradesh and North Eastern states are the major flower growing states in India. Andhra Pradesh is having largest area *i.e.* 34.85 ('000'ha) of flowers. Tamil Nadu is the largest loose flower producing state *i.e.* 312.97 ('000'Mt), while West Bengal is the leading cut flower producing state *i.e.* 25429.1 (lakh Nos) in India (Anonymous, 2013). While ornamental plants are produced mainly for their aesthetic value, the propagation and improvement of quality attributes such as leaf types, flower colour and fragrance, longevity and form and the creation of novel variation are the important economic goals for the expanding ornamental industry (Verma *et al.*, 2011). Loose flower uses for making garlands, veni and bracelets as well as for worshipping (Bohra and Kumar, 2014). Organic farming may be a desirable proposition for improving the quality of horticultural produce and soil health; it is difficult to meet the nutrient requirement of the crops, exclusively through organic farming. The use of organic manures and bio-fertilizers along with the balance use of chemical fertilizers is known to improve physico-chemical and biological properties of soil, besides improving the efficiency of applied chemical fertilizers (Patanwar *et al.*, 2014). In addition, they are ecofriendly, easily available and cost-effective. In many areas of the country, the overall health and productivity of the soil have declined due to use of high yielding varieties and enhanced application of chemical fertilizers. High yielding varieties of crops can perform to their potential, only if they are grown in productive soils. Nutrient status of the plants can be pointer to the response of plant to the fertilization and internal content of the nutrient determine the fertilizer requirements (Polar *et al.*, 2014). The quality of flowers is greatly influenced by the quantity and source of nutrients. At present, these nutrients are primarily supplied through chemical fertilizers. Therefore, recently the use of organic manures *viz.*, farm yard manure, vermicompost and biofertilizers like Azospirillum and Phosphate Solubilizer Bacteria and Phosphate Solubilizer microorganism is gaining momentum. Therefore, under these circumstances, our objective is to evolve judicious combination of organic manures, biofertilizers and chemical fertilizers, it may be feasible and viable option for sustainable production on a commercial and profitable scale and is the need of hour.

Organic manure

The practical way to improve the quality of soil which a low organic matter content is of the addition of organic materials to the soil either fresh or composted (Garcia *et al.*, 2000 and Pascual *et al.*, 1999). These materials are rich in labile carbon function which acts as a source of energy for microorganisms also increasing soil microbial populations and their activities (Papavizas *et al.*, 1982). Organic materials also increase soil water holding capacity and aggregation and improve the nutrient status (Ros *et al.*, 2001). In addition organic matter is a necessary food for the soil organisms. The combination of organic matter like farm yard manure and mineral

*Corresponding author

fertilizers provides the ideal environmental conditions. The use of manures as an organic source occupy an important place as they provide a scope for reduction on use of costly chemical fertilizers which can pollute soil in long term use (Sharma, 2005). The effect of FYM on number of flowers can also be attributed to the fact that there will be higher translocation of carbohydrates from other parts to reproductive parts during development. The source to sink translocation can be more effective only when the rate of photosynthesis is high (Johnson, 1984). Srinivas and Gowda, (1999) reported in China aster that the application of FYM at the rate of 15 t/ha along with recommended NPK increased in number of flowers/plant and flower yield. In chrysanthemum that application of FYM resulted in better growth and yield like taller plant and more number of leaves, number of flowers plant⁻¹, large size flower and yield plant⁻¹. Similar results were found by Haripriya and Sriramachandrasekharan, (2002) in chrysanthemum and Prakash *et al.*, (2002) in marigold.

Vermicomposts are products derived from the accelerated biological degradation of organic wastes by earthworms and microorganisms (Edwards *et al.*, 1998). Vermicompost besides being a rich source of micronutrients, also act as chelating agent and regulates the availability of metabolic micronutrients like iron and zinc to the plants and increases the plant growth and yield by providing nutrients in the available form. Use of vermicompost in agriculture was first reported by (Hopp and Slater, 1979). Further, they quantified the response of crop to earthworms in terms of yield. Vermicompost serves as organic manure, since it is a source of nutrients viz., nitrogen, phosphorus, potassium, humic acids and micronutrients (Biradar and Shaila, 2006). Maximum plant height, number of leaves, number of flowers plant⁻¹ and flower yield hectare⁻¹ due to the application of vermicompost 10 tonnes per hectare and recommended dose of NPK in China aster (Nethra, 1996, Chaitra and Patil, 2007, Sonawane *et al.*, 2009, Gharat *et al.*, 2008 and Balajiet *et al.*, 2006). In gladiolus yield and quality is increased by the application of vermicompost (Gangadharan and Gopi-nath, 2000). In chrysanthemum plant grown in 50% vermicompost had a greater growth index at harvest, number of flowers plant⁻¹ as well as early flower development the result are in line with Angadi, (2010), Hidalgo and Harkers, (2002) in chrysanthemum and Bashan, (2004). In marigold application of vermicompost (60%) resulted in the highest weight, size and dry weight of shoot, but the maximum bush height was obtained by 20% vermicompost. The highest number of lateral branches was recorded in 40% vermicompost treatment. Similar results were found by Atiyeh *et al.* (2000), Chauhan *et al.* (2005) in calendula, Patel *et al.* (2011), Shadanpour *et al.* (2011) and Ajitkumar, (2002) in marigold.

Bio-fertilizers

Bio-fertilizers are biological preparations of live or latent cell of selected strain of nitrogen fixing or phosphorous solubilizing

beneficial soil microorganisms (Bhaskaran *et al.*, 2002). These may help in mineralization of nitrogen and transformation of several elements like sulphur and iron into available forms. These biofertilizers benefit flower production by making the nutrients available to plants. Common biofertilizers used in horticultural crops are Azospirillum, Azotobacter and PSB. Azospirillum is a non-symbiotic, N fixing bacterium, which fixes the atmospheric nitrogen when inoculated to plants and helps in saving the application of N fertilizers to an extent of 20-25 percent (Subbarao, 1993). Wild strain of Azospirillum fixes atmospheric nitrogen efficiently either as a free living bacterium or in combination with plants and thus participates in the nitrogen cycle (Tarrand *et al.*, 1978). It is associated with root tissues and lives in harmony with root tissues, fixing substantial amount of nitrogen. Increased growth in marigold by inoculation of Azospirillum, this might be due to added nitrogen to crop through associative symbiosis and increased production of growth hormones like NAA, GA and cytokinins (Bashan, 2004 and Balasubramanian, 1989). (Preethi, 1990) Early flowering in rose by the inoculation of Azospirillum might be due to induced cytokinin synthesis and rapid assimilation of photosynthates resulting in early transformation of the axillary bud from vegetative to reproductive phase (Bashan, 2004). Wange *et al.* (1995) reported that increase in number of bulblets (14.3/plant), flower stalks (23348 dozen/ha) with higher monetary returns (1.00:3.73) in tuberose Cv. 'Single' by the inoculation of Azospirillum. Increase in vegetative growth in China aster by bio-fertilizers was because of stimulation in nutrient uptake and biosynthesis of plant growth regulators similar result were found by Prabhatkumaret *et al.*, (2003).

subsequent years several other types of Azotobacteria group have been found in the soil and rhizosphere such as Azotobacter vinelandii, (Lipman, 1903), Azotobacter beijerinckii, (Lipman, 1904), Azotobacter nigricans, (Krassilnikov, 1949), Azotobacter paspali, (Döbereiner, 1966, Thompson and Skerman 1980) Azotobacter armenicus (Thompson and Skerman, 1981), Azotobacter salinestris, (Page and Shivprasad, 1991). Three possible mechanisms have been proposed: N₂ fixation, delivering combined nitrogen to the plant, the production of phytohormone like substances that alter plant growth and morphology and bacterial nitrate reduction which increases nitrogen accumulation in inoculated plants (Triplett 1996). Biological nitrogen fixation is one way of converting elemental nitrogen into plant usable form (Gothwalet *et al.*, 2007). Nitrogen-fixing bacteria (NFB) that function transform inert atmospheric N₂ to organic compounds (Bakulinet *et al.*, 2007). These nitrogen-fixing bacteria are important for ecology and agriculture. Maximum leaf area, length of first order lateral shoots, number and weight of flowers/m² during first and second flush were observed with inoculation of Azotobacter in rose (Singh, 2005). Azotobacter when applied in marigold with Azospirillum under different level of chemical nitrogen increase in growth, yield and biochemical attributes (i.e.

Table 1: Nutrients present in vermicompost

N (%)	P ₂ O ₅ (%)	K ₂ O (%)	Cu (ppm)	Mn (ppm)	Fe (ppm)	Zn (ppm)	Ca (ppm)	Mg (ppm)	S (ppm)
0.9-1.5	0.3-0.64	0.54-1.72	2.00-37.7	10.0-105.0	10.0-161.00	5.7-120.0	1.0-1.75	0.47-1.75	0.50-1.50

[Sources: Kale, (1998), Chaudhuri *et al.*, (2000), Kittumathet *et al.*, (2007), Giraddi, (2007) and (2011), Meenatchi, (2008) and Waseem *et al.*, (2013)]

chlorophyll, total sugar, total free amino acid, and nitrate reductase). Phosphorus is a major growth limiting nutrient and unlike the case of nitrogen, there is no large atmospheric source that can be made biologically available (Ezawa *et al.* 2002). Root development, stalk and stem strength, flower and seed formation, crop maturity and production, crop quality and resistance to plant diseases are the attributes associated with phosphorus nutrition. Higher crop yields result from solubilization of fixed soil P and applied phosphates by PSB (Zaidi (1999). Microorganisms with phosphate solubilizing potential increase the availability of soluble phosphate and enhance the plant growth by improving biological nitrogen fixation (Kuceyet *al.*, 1989; Ponnurugan and Gopi, 2006). Bacteria are more effective in phosphorus solubilization than fungi (Alamet *al.*, 2002). Among the whole microbial population in soil, PSB constitute 1 to 50 % (Chen *et al.*, 2006). Number of PSB among total PSM in north Iranian soil was around 88 % (Fallah, 2006). Among the soil bacterial communities, ectorhizospheric strains from *Pseudomonas* and *Bacilli*, and endosymbiotic rhizobia have been described as effective phosphate solubilizers (Igualet *al.*, 2001). Strains from bacterial genera *Pseudomonas*, *Bacillus*, *Rhizobium* and *Enterobacter* along with *Penicillium* and *Aspergillus* fungi are

Table 2: Microbial population in vermicompost

Bacteria (cfu/g)	Fungi (cfu/g)	Actinomycetes (cfu/g)
5×10^6 to 10×10^7	10×10^3 to 27×10^3	2.2×10^4 to 15×10^7

[Sources: Jones *et al.*, (2005), Giraddi, (2007) and (2011), Meenatchiet *al.*, (2009) and Waseem *et al.*, (2013)]

Table 3: INM treatments on chrysanthemum and marigold flowers

S. No.	Treatments	Chrysanthemum Treatment detail	Marigold Treatment detail
1.	T ₁	100% RDF	100% RDF
2.	T ₂	50% RD'N' through FYM + 50% RDF	50% VC equivalent to RD'N' + 50% RDF
3.	T ₃	50% RD'N' through VC + 50% RDF	Azo + 75% RD'N' + 100% RD'P' and 'K'
4.	T ₄	Azo + 75% RD'N' + 100% RD'P' and RD'K'	PSB + 75% RD'P' + 100% RD'N' and 'K'
5.	T ₅	PSB + 75% RD'P' + 100% RD'N' and RD'K'	Cow Urine (5%) + 75% RD'N' + 100% RD'P' and 'K'
6.	T ₆	Azo + 50% RD'N' through FYM + 50% RDF	Azo + 50% VC equivalent to RD'N' + 50% RDF
7.	T ₇	Azo + 50% RD'N' through VC + 50% RDF	PSB + 50% VC equivalent to RD'N' + 50% RDF
8.	T ₈	PSB + 50% RD'N' through FYM + 50% RDF	Cow Urine (5%) + 50% VC equivalent to RD'N' + 50% RDF
9.	T ₉	PSB + 50% RD'N' through VC + 50% RDF	Azo + PSB + 50% RD'N' and 'P' + 100% RD'K'
10.	T ₁₀	Azo + PSB + 50% RD'N' and 'P' + 100% RD'K'	Azo + PSB + Cow Urine (5%) + 50% RD'N' and 'P' + 100% RD'K'
11.	T ₁₁	Azo + PSB + 50% RD'N' through FYM + 50% RDF	Azo + PSB + 50% VC equivalent to RD'N' + 50% RDF
12.	T ₁₂	Azo + PSB + 50% RD'N' through VC + 50% RDF	Azo + PSB + Cow Urine (5%) + 50% VC equivalent to RD'N' + 50% RDF

(source-Patanwar *et al.*, 2014, Sahu, 2014)

Table 4: Phosphorus solubilization potential of various bacterial strains at different pH values of the medium

PSB strains	Estimated quantity in a standing liquid culture		References
	P (µg mL ⁻¹)	Ph	
D 5/23 Pantoea agglomerans	62.76	5.93	Amellalet <i>al.</i> , (1999).
CC 322 Azospirillum sp. Mac 27	83.39	6.19	Rattiet <i>al.</i> , (2001).
Azotobacter chroococcum Ala 27	98.11	4.84	Narulaet <i>al.</i> , (2000).
Azotobacter chroococcum Msx 9	1.10	7.50	Narulaet <i>al.</i> , (2000).
Azotobacter chroococcum ER 3	65.90	5.82	Narulaet <i>al.</i> , (2000).

the most powerful P solubilizers (Whitelaw, 2000). Kumar *et al.* (2006) observed in Marigold cv. Pusa Narangi that FYM + PSB followed by FYM + Azotobacter resulted in maximum yield per plant, flower diameter, average flower weight and fresh weight of flower per plant. In dahlia height of plant, number of leaves per plant, plant spread, earliness of flowering and yield of flower was superior in the plants receiving vermicompost (500 g) with PSB 25 g per plot (Waradeet *al.* 2007). In China aster application of full dose of nitrogen in addition of Azotobacter and PSB resulted in more plant weight, stem diameter, number of branches, number of leaves, while, flower yield was maximum under Azotobacter and PSB with 75% N (Kulkarni *et al.*, 1996).

Inorganic fertilizer

The application of chemical fertilizer in balanced dose helpful in increase the production of flowers as nitrogen is a component of amino acid, which is essential for protein synthesis. Nitrogen bases like pyrimidines and purines and many co-enzymes need nitrogen for their synthesis. It is also a component of cytochrome and chlorophyll which are essential for photosynthesis. While Phosphorus forms a source of energy in the form of ATP and ADP, the cell division is also influenced by phosphorus. It is also a component of many enzymes, co-enzymes, nucleic acids and phospholipids and potash acts as catalyst for various enzymes and co-enzymes and starch synthesis takes place in the presence of potash. It has a vital role in photo respiration, translocation of metabolites and transpiration. In annual chrysanthemum

cv. Nivea, plant height and number of primary branches per



Figure 1:

plant at full bloom were at maximum by the application of nitrogen at 300 kg ha⁻¹, phosphorus at 200 kg ha⁻¹ and potash 160 kg ha⁻¹. Similar results were found by Baboo and Sharma, (1997), Dixit et al. (2004) and Dorajeerao et al. (2012). In marigold cv. African giant, Effect of N:P:K on flowering attributes was highest number of flowers per plant, number of seeds per flower, seed yield per plant and plant germination percentage. The results are also corroborated with Gavanheet al. (2004), Acharya and Dashora, (2004), Kore et al. (2004), Naik et al. (2005), Karuppaiah and Krishna, (2005), Das et al. (2005) and Jawaharlal et al. (2013). (Gurav et al., 2004) In gerbera found that application of nitrogen @ 15 g/m²/month significantly produced more number of flowers per plant (29.88), flowers with longer stalk length (47.33 cm) and larger diameter (8.75 cm). Application of phosphorus @ 20 g/m²/month significantly produced more number of flowers per plant (29.55) and flowers with larger diameter (8.70 cm). In *Calendula officinalis* effect of nitrogen fertilizer was significant on dried and fresh weight of flower and that the highest one obtained from 90 kg of N ha⁻¹ treatment reported by Mollafilabi et al. (2013).

REFERENCES

- Acharya, M. M. and Dashora, L. K. 2004. Response of graded levels of nitrogen and phosphorus on vegetative growth and flowering in African marigold (*Tagetes erecta* L.). *J. Orn. Hort.* **7(2)**: 179-183.
- Ajitkumar 2002. Effect of organic and inorganic fertilizers on growth, yield and post harvest life of marigold. *M. Sc. (Ag.) Thesis*, UAS, Dharwad.
- Alam, S., Khalil, S., Ayub, N. and Rashid, M. 2002. *In vitro* solubilization of inorganic phosphate by phosphate solubilizing microorganism (PSM) from maize rhizosphere. *Intl. J. Agri. Bio.* **4**: 454-458.
- Amellal, N., Bartoli, F., Villemin, G., Talouizte, A. and Heulin, T. 1999. Effects of inoculation of EPS-producing *Pantoea agglomerans* on wheat rhizosphere aggregation. *Plant Soil.* **211**: 93-101.
- Angadi, P. 2010. Integrated nutrient management studies on growth, yield and quality of garland chrysanthemum (*Chrysanthemum coronarium* L.). *M. Sc. (Ag.) Thesis*, UAS, Dharwad.
- Anonymous. 2013. Report of national horticulture board, Indian horticulture database. p. 283.
- Atiyeh, R. M., Edwards, C. A., Subler, S. and Metzger, J. D. 2000. Earthworm processed organic waste as components of horticulture potting media for growing marigold and vegetable seedlings. *Compost Sci. Util.* **8**: 215-223.
- Baboo, R. and Sharma, K. S. K. 1997. Effect of nitrogen and potassium fertilization on growth and flowering of annual chrysanthemum. *J. Orn. Hort.* **5(1-2)**: 44-45.
- Balaji, S. K., Reddy, B. S., Patil, B. C. and Divakara, A. 2006. Influence of vermicompost and *in situ* vermiculture on the quality attributes and saleable yield in China aster. *Scientia Hort.* **10**: 217-221.
- Balasubramanian, J. 1989. Studies on the combined effect of *Azospirillum*, vesicular arbuscular mycorrhizal and inorganic fertilizers on growth performance of French marigold (*Tagetesputula* L.). *M. Sc. (Agri.) Thesis*, TNAU., Coimbatore.
- Bakulin, M. K., Grudtsyna, A. S. and Pletneva, A. 2007. Biological fixation of nitrogen and growth of bacteria of the genus *Azotobacter* in liquid media in the presence of Perfluoro carbons. *Appl. Bioch. Microb.* **4**: 399-402.
- Bashan, Y. and Holguin, G. 2004. *Azospirillum*-plant relationships: physiological, molecular, agricultural, and environmental advances (1997-2003). *Can. J. Microb.* **50(8)**: 521-577.
- Beijerinck, M. W. 1901. Überlogonitro philemikroben. *Zentralbl. Bakteriol. Parasitenkd. Infektionskr. II Abt.* pp. 561-582.
- Bhaskaran, P. A. and Jayabalan, N. 2002. Usefulness of biofertilizers in economizing nitrogenous fertilizers in *Tagetes erecta* L. *J. Phyto. Res.* **15(2)**: 155-160.
- Biradar, A. P. and Shaila, H. M. 2006. Studies on suitability of vermicomposting structures. *Karnataka. J. Agri. Sci.* **19**: 411-413.
- Bohra, M. and Kumar, A. 2014. Studies on effect of organic manure and bioinoculants on vegetative and floral attributes of chrysanthemum cv. Little darling. *The Bioscan.* **9(3)**: 1007-1010.
- Chaudhuri, P. S., Bhattacharjee, G. and Dey, S. K. 2000. Chemical changes during vermicomposting (*Perionyx excavatus*) of Kitchen waste. *Tropical Ecol.* **41**: 107-110.
- Chaitra, R. and Patil, V. S. 2007. Integrated nutrient management studies in china aster (*Callistephus chinensis* Nees) cv. Kamini. *K.J. Agri. Sci.* **20(3)**: 689-690.
- Chauhan, A. and Kumar, V. 2007. Effect of graded levels of nitrogen and VAM on growth and flowering in calendula (*Calendula officinalis* L.). *Ind. J. Hort.* **10(1)**: 61-63.
- Chen, Y. P., Rekha, P. D., Arunshen, A. B., Lai, W. A. and Young, C. C. 2006. Phosphate solubilizing bacteria from subtropical soil and their tri-calcium phosphate solubilizing abilities. *App. Soil Ecol.* **34**:

33-41.

- Das, J. N. and Mishra, H. N. 2005.** Studies on graded doses of fertilizers and polythene mulches on growth, flowering and yield of African marigold (*Tagetes erecta* L.) cv. Siracole. *Ori. J. Hort.* **33(2)**: 42-45.
- Dixit, A., Trivedi, J. and Verma, S. 2004.** Effect of nitrogen and phosphorus on growth yield and quality of chrysanthemum cv. Local white under Chhattisgarh region. *Plant Arch.* **4(1)**: 171-173.
- Döberainer J. 1966.** *Azotobacter paspalis* sp. nov., uma bacteria fixadora de nitrogenio na rizosfera de Paspalum. *Pesquisa Agropecuaria Brasileira*. **1**: 357-365.
- Dorajeerao, A. V. D., Mokashi, A. N., Patil, V. S., Venugopal, C. K., Lingaraju, S. and Koti, R. V. 2012.** Effect of graded levels of nitrogen and phosphorus on growth and yield of garland chrysanthemum (*Chrysanthemum coronarium* L.). *K. J. Agri. Sci.* **25(2)**: 224-228.
- Edwards, C. A. and Fletcher, K. E. 1998.** Interactions between earthworms and microorganisms in organic matter breakdown. *Agri. Eco. Environ.* **24**: 225-247.
- Ezawa, T., Smith, S. E. and Smith, F. A. 2002.** P metabolism and transport in AM fungi. *Plant Soil.* **244**: 221-230.
- Fallah, A. 2006.** Abundance and distribution of phosphate solubilizing bacteria and fungi in some soil samples from north of Iran. 18th World Congress of Soil Science, July 9-15, Philadelphia, Pennsylvania, USA.
- Gangadharan, G. D. and Gopinath, G. 2000.** Effect of organic and inorganic fertilizers on growth, flowering and quality of gladiolus Cv. white prosperity. *K. J. Agri. Sci.* **11(3)**: 401-405.
- Garcia, C., Hernandez, T., Roldan, A., Albaladejo, J. and Castillo, V. 2000.** Organic amendment and mycorrhizal inoculation as a practice in afforestation of soils with pinus halepensis miller, effect on their microbial activity. *Soil Bio. Bioch.* **16**: 1173-1181.
- Gavhane, P. B., Kore, V. N., Dixit, A. J. and Gondhali, B. V. 2004.** Effect of graded doses of fertilizers and polythene mulches on growth, flower quality and yield of marigold (*Tagetes erecta* L.) Cv. Pusa Narangi Gainda. *Ori. J. Hort.* **32(1)**: 35-37.
- Gharat, S. N., Rohidas, S. B., Patil, M. B. and Nawghare, P. D. 2008.** Impact of organic and inorganic fertilizers on yield and quality parameters of china aster flower var. California giant. *J. MH. Agri. Uni.* **33(1)**: 103-104.
- Giraddi, R. S. 2007.** Vermitechnologies (in Kannada), UAS, Dharwad, India, p. 62.
- Giraddi, R. S. 2011.** Research priorities in vermitechnologies. Final report submitted to NABARD, Mumbai, p.106.
- Gothwal, R. K., Nigam, V. K., Mohan, M. K., Sasmal, D. and Ghosh, P. 2007.** Screening of nitrogen fixers from rhizospheric bacterial isolates associated with important desert plants. *App. Eco. Environ. Res.* **6(2)**: 101-109.
- Gurav, S. B., Katwate, S. M., Singh, B. R., Sabale, R. N., Kakade, D. S. and Dhane, A. V. 2004.** Effect of nutritional levels on yield and quality of gerbera. *J. Orn. Hort.* **7(3-4)**: 226-229.
- Haripriya, K. and Sriramachandrasekharan, M. V. 2002.** Effect of organic amendments on the growth and yield of chrysanthemum. *J. Eco.* **14(1)**: 39-42.
- Hidalgo, P. R. and Harkers, R. L. 2002.** Earthworms eating as a substrate amendment for chrysanthemum production. *Hort. Sci.* **37(7)**: 1035-1039.
- Hopp, H. and Slatter, C. S. 1979.** The effect of earth worms on the productivity of agricultural soil. *J. Agri. Res.* **78**: 325-329.
- Igual, J. M., Valverde, A., Cervantes, E. and Velázquez, E. 2001.** Phosphate-solubilizing bacteria as inoculants for agriculture: use of updated molecular techniques in their study. *Agronomie.* **21**: 561-568.
- Jawaharlal, M., Swapna, C. and Ganga, M. 2013.** Comparative analysis of conventional and precision farming systems for African marigold (*Tagetes erecta* L.). *Acta. Hort.* **970**: 311-318.
- Johnson, C. R. 1984.** Phosphorus nutrition on mycorrhizal colonization, photosynthesis, growth and nutrient composition of *Citrus aurantium*. *Plant Soil.* **80**: 35-42.
- Jones, P. N., Biradar, A. P., Patil, M. B. 2005.** Microflora associated with vermicompost obtained from different weeds. *K. J. Agri. Sci.* **18**: 186-187.
- Kale, R. D. 1998.** Earthworm-Cinderella of Organic Farming, Prism Books Pvt. Ltd., Bangalore. p. 87.
- Karuppaiah, P. and Krishna, G. 2005.** Response of spacing and nitrogen levels on growth, flowering and yield characters of French marigold (*Tagetes patula* L.). *Ind. J. Hort.* **8(2)**: 96-99.
- Kitturmath, M. S., Giraddi, R. S. and Basavaraj, B. 2007.** Nutrient changes during earthworm, *Eudriluseugeniae* (Kinberg) mediated vermicomposting of agro industrial wastes. *K. J. Agri. Sci.* **20**: 653-654.
- Kore, V. N., Gavhane, P. B., Patil, R., Gondhali, B. V. and Bendale, V. W. 2004.** Effect of graded levels of fertilizers and polythene mulches on growth and yield of marigold. *J. Soils Crops.* **14(2)**: 258-261.
- Krassilnikov, N. A. 1949.** Opredeliteli Bakterii Aktinomycetov. Izd. AN sssr. M.
- Kucey, R. M. N., Janzen, H. H. and Legget, M. E. 1989.** Microbial mediated increases in plant available phosphorus. *Adv. Agron.* **42**: 199-228.
- Kulkarni, B. S., Nalawadi, U. G. and Giraddi, R. S. 1996.** Effect of vermicompost and vermiculture on growth and yield of china aster (*Callistephus chinensis* Nees) cv. Ostrich plum mixed. *S. Ind. Hort.* **44(1&2)**: 33-35.
- Kumar, M., Sharma, S. K., Singh, S., Dahiya, D. S., Mohammed, S. and Singh, V. P. 2006.** Effect of farm yard manure and different biofertilizers on yield and nutrient content of marigold cv. Pusa Narangi. *Har. J. Hort. Sci.* **35(3-4)**: 256-257.
- Lipman, J. G. 1903.** Report on the New Jersey Agricultural Experiment Station. **24**: 217-285.
- Lipman, J. G. 1904.** Report on the New Jersey Agricultural Experiment Station. **25**: 237-289.
- Madhuri, D. 2011.** Discemination of vermitechnology to Women. *DST (GOI) Report*, New Delhi. p. 45.
- Meenatchi, R. 2008.** Molecular characterization of earthworms, nutrient assessment and use of vermitechnologies in pest management. *Ph. D. Thesis*, UAS, Dharwad, Karnataka.
- Meenatchi, R., Giraddi, R.S. and Biradar, D.P. 2009.** Assessment of genetic variability among strains of earthworm, *Eudriluseugeniae* (Kinberg) using PCR RAP D technique. *K. J. Agri. Sci.* **22**: 942-945.
- Mollafilabi, A. and Hosseini, H. 2013.** Effect of different seed rates and nitrogen fertilizer on yield and yield components of calendula (*Calendula officinalis*). *Acta Hort.* **997**: 43-50.
- Patanwar, M., Sharma, G., Banjare, C., Chandravanshi, D. and Sahu, E. 2014.** Growth and development of chrysanthemum (*dendranthema grandifloratzelev*) as influenced by integrated nutrient management. *The ecoscan.* **6**: 459-462.
- Naik, B. H., Patil, A. A., Patil, V. S. and Basavaraj, N. 2005.** Effect of nitrogen and phosphorus on xanthophyll yield of marigold. *K. J. Agri. Sci.* **1(3)**: 81-88.
- Narula, N., Kumar, V. R. K., Behl, Deubel, A., Gransee, A. and Merbach, W. 2000.** Effect of P-solubilizing *Azotobacter chroococcum* N, P, K uptake in P-responsive wheat genotypes grown under greenhouse conditions. *J. Plant Nutri. Soil Sci.* **163**: 393-398.
- Nethra, N. N. 1996.** Effect of organic and inorganic fertilizers on

- growth, yield and postharvest life of china aster (*Callistephus chinensis* Nees.). *M. Sc. (Hort.) Thesis*, UAS, Bangalore.
- Page, W.J. and Shivprasad, S. 1991.** *Azotobacter salinestrissp. nov.*, a sodium-dependent, microaerophilic, and aeroadaptive nitrogen-fixing bacterium. *Int. J. Sys. Bacteriol.* **41**: 369-376.
- Papavizas, G. C. and Lumsden, R. D. 1982.** Improved medium for isolation of *Trichoderma spp.* From soil. *Plant Dis.* **66**: 1019-1020.
- Pascual, J. A., Garcia, C and Hernandez, T. 1999.** Comparison of fresh and composted organic waste in their efficacy for the improvement of arid soil quality. *Biores. Tech.* **68**: 255-264.
- Patel, P. R., Patel, N. K., Valia, R. Z. and Chaudhari, S. R. 2011.** Effect of nitrogen and vermicompost on floral and yield parameters of african marigold. *Asi. J. Hort.* **6(2)**: 478-480.
- Polara, N. D., Gajipara, N. N. and Barad, A. V. 2014.** Effect of nitrogen and phosphorous on nutrient content and uptake in different varieties of African marigold (*Tagetes erecta* L.). *The Bioscan.* **9(1)**: 115-119.
- Ponmurugan, P. and Gopi, C. 2006.** Distribution pattern and screening of phosphate solubilising bacteria isolated from different food and forage crops. *J. Agron.* **5**: 600-604.
- Prabhatkumar, S., Raghava, P. S. and Mishra, R. L. 2003.** Effect of biofertilizers on growth and yield of China Aster. *J. Orn. Hort.* **6(2)**: 85-88.
- Prakash, A., Sharma, S. K., Sindhu, S. S. and Prakash, A. 2002.** Effect of phosphorous and FYM on NPK content of marigold in chloride dominated soil. *Har. J.Hort. Sci.* **31(1&2)**: 47-49.
- Preethi, T. L. 1990.** Studies on the effect of nitrogen, *Azospirillum* and ascorbic acid on growth and flowering of Edward rose (*Rosa bourboniana* Desp.). *M. Sc. (Agri.) Thesis*, TNAU, Coimbatore.
- Ratti, N., Kumar, S., Verma, H. N. and Gautam, S. P. 2001.** Improvement in bioavailability of tricalcium phosphate to *Cymbopogon martinii* var. *motiaby* rhizobacteria, AMF and *Azospirillum* inoculation. *Microb. Res.* **156**: 145-149.
- Ros, M., Garcia, C. and Hernandez, T. 2001.** The use of organic waste in the control of erosion in a semi-arid mediterranean soil. *Soil Use Manag.* **17**: 292-293.
- Sahu, N. P. 2014.** Effect of bio-organic and inorganic nutrient sources on the growth and flower yield of marigold. *M. Sc. (Agri.) Thesis*, IGKV, Raipur.
- Shadanpour, F., Torkashvand, A. M. and Majd, K. H. 2011.** Marigold the possibility using vermicompost as the growth medium. *J. Orn. Hort. Plants.* **1(3)**: 153-160.
- Sharma, A. K. 2005.** The living soil Bio-fertilizers for Sustainable Agriculture, Agrobios (India), Jodhpur, pp. 1-19.
- Singh, A. K. 2005.** Response of rose plant growth and flowering to nitrogen, *Azotobacter* and farm yard manure. *J. Orn. Hort.* **8(4)**: 296-298.
- Srinivasa, K. N. and Narayana Gowda, J. V. 1999.** Effect of different organic manures on growth and flower yield of china aster (*Callistephus chinensis* (L.) Nees.). *Crop Res.* **18**: 104-107.
- Sonawane, S. P., Dabke, D. J., Dabke, S. B. and Dhane, S. S. 2009.** Effect of nitrogen, and FYM on yield and nutrient uptake by china aster (*Callistephus chinensis* (L.) Nees.). *J. Mh. Agri. Uni.* **34(1)**: 90-91.
- Subbarao, N. S., Venkataraman, G. S. and Kannaiyan, S. 1993.** Biological nitrogen fixation. *Allied publication*, 104 A Mayapuri, New Delhi.
- Tarrand, J. J., Krieg, N. R. and Doberreiner, J. 1978.** A taxonomic study of the *Spirillum lipoferum* group, with description of new genus *Azospirillum* gen. nov. and species *Azospirillum lipoferum* (Bejerink) Comb. nov. and *Azospirillum brasiliense* Sp. nov. *Can. J. Micro.* **27**: 967-980.
- Thompson, J.P. and Skerman V.B.D. 1980.** *Azotobacteraceae*: the taxonomy and ecology of the aerobic nitrogen fixing bacteria. Academic Press, London.
- Thompson, J.P. and Skerman V.B.D. 1981.** Validation list No6. *Int. J. Syst. Bacteriol.*, **31**: 215-218.
- Triplet, E.W. 1996.** Diazotrophic endophytes: progress and prospects for nitrogen fixation in monocots. *Plant Soil* **186**: 29-38.
- Verma, S. K., Angadi, S. G., Patil, V. S., Mokashi, A. N., Mathad, J. C. and Mummigatti, U.V. 2011.** Growth, yield and quality of chrysanthemum (*Chrysanthemum morifolium* Ramat.) cv. Raja as influenced by integrated nutrient management. *K. J. Agri. Sci.* **24(5)**: 681-683.
- Wange, S.S., Patil, P.L. and Patil, J.J. 1995.** Effect of biofertilizers alone and with nitrogen levels on tuberose cv. Single petated. *J. Soils and Crops* **5(2)**: 97-99.
- Warade, A. P., Golliwar, V. J., Chopde, N., Lanje, P. W. and Thakre, S. A. 2007.** Effect of organic manures and biofertilizers on growth, flowering and yield of dahlia. *J. Soils Crops* **17(2)**: 354-357.
- Waseem, M. A., Giraddi, R. S. and Math, K. K. 2013.** Assessment of nutrients and micro flora in vermicompost enriched with various organics. *J. Exp Zoo* **116**: 697-703.
- Whitelaw, M. A. 2000.** Growth promotion of plants inoculated with phosphate solubilizing fungi. *Adv. Agron.* **69**: 99-151.
- Zaidi, A. 1999.** Synergistic interactions of nitrogen fixing microorganisms with phosphate mobilizing microorganisms. Ph.D. Thesis, Aligarh Muslim University, Aligarh, India.

